

Molecular and Morphological Identification of *Balamuthia mandrillaris* (Free- living Amoeba) Isolated From Environmental Sources in Thi- Qar province / Iraq

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Abstract

Balamuthia mandrillaris is a free- living amoeba that lives in soil and water near human settlements . It was supposed to occupy the same ecological habitats of *Acanthamoeba* . *B. mandrillaris* has been reported as the causative agent of fatal *Balamuthia* Amoebic Encephalitis (BAE) essentially, there is a little information about this genus. So that, this study attempted to determine characterization of *B. mandrillaris* at molecular level and morphological features. Seventy five of environmental samples were collected from different sources in Thi-Qar province south Iraq during the period February to September 2020. Samples were screened for *B. mandrillaris* and identified by morphologically and genetically by PCR method. Results stated that *B. mandrillaris* cyst and trophozoite were observed in some environmental sample were 12 (16%). There was only 9 (12%) were positive out of 12 suspected environmental isolates that confirmed by the presence of distinct amoeba cyst or both trophozoites and cyst by using PCR method including *Balamuthia mandrillaris* specific primer Balspec 16S forward and reverse that amplified a portion of mitochondrial rRNA gene yield a 1075 bp product . Trophozoite and cyst were observed in culture ,the trophozoite stage approximately 35-50 micron in diameter. it has finger like pseudopodia the trophozoite showed an ability to produce pseudopodia from any part of the amoebic body , the cysts of *B. mandrillaris* were spherical and approximately 12-25 micron surrounded by a double layer membrane , an inner wall that was endocyst and an outer wrinkled wall that was ectocyst. the ectocyst had no pores like *Acanthamoeba* and endocyst without arms . The presence of *B. mandrillaris* can be considered a health hazard , as *B. mandrillaris* is an opportunistic amoeba and also may harbor some pathogenic bacteria .

Keywords : *Balamuthia mandrillaris* , free-living amoeba , Thi-Qar province , Iraq , opportunistic amoeba

INTRODUCTION

Balamuthia mandrillaris is a protista pathogen that can cause encephalitis with a fatality rate of > 95% (Siddiqui and Khan , 2015) . *B. mandrillaris* is a free – living amoeba found in the

environment causes rare cases of human disease including cutaneous and central nervous system (CNS) disease called granulomatous amebic encephalitis (GAE) (Jennifer *et al.*, 2019). The disease it causes are similar to those of *Acanthamoeba* spp., including granulomatous amebic encephalitis and cutaneous lesions (Visvsevara *et al.*, 2007).

B. mandrillaris is the only species of *Balamuthia* known to cause infection in humans or animals, *Balamuthia* was first isolated from the brain of a mandrill baboon that died at the San Diego Zoo from meningoencephalitis. The amoebae was first described as a leptomyxid amoebae but later identified and named *Balamuthia mandrillaris* (Schuster and Visvsevara, 2008).

B. mandrillaris had ability to host bacteria, enhances their infective capacity for host cells or support transmission to a new susceptible hosts, thus amoeba may serve as a biological host as well as a transmission vector (Matin *et al.*, 2008).

B. mandrillaris isolation and to cultivation is difficult (Schuster, 2002), but thought to be ubiquitous in the environment (MMWR, 2010). The first environment isolation of the amoeba was from soil by Lokhande *et al.* (2015), it was also recorded from two dogs who swam in pond water previously (Finnine *et al.*, 2007).

MATERIAL AND METHODS :

Sample collection & cultivation:

Samples were collected from different environmental source, including soil, water from (rivers, tapwater, the marshes, ponds, drops of water from the air conditioning equipment outside the building), and animals wastes, these samples collected from different region in Thi-Qar province during the period from February to September 2020.

A- Water samples : water samples were collected in 100 ml sterile cups, the date and site details were fixed for each sample. In the lab. 3-5 ml of each sample was cultured on non-nutrient agar (NN-agar) medium in two replicates within 24 hours of collection incubated in 26 C⁰ and amoebic growth was examined daily by light microscope on slide or inverted microscope on agar and followed for 4 weeks.

B- Soil samples : soil samples and animal wastes were collected in sterile containers the date and site details were fixed for each sample, within the next 24 hours of collection from each sample 3 grams were suspended in 5 ml of sterile distilled water and supernatant was cultured on non-nutrient agar (NN-agar) medium in two replicates within 24 hours of collection incubated in 26 C⁰ with 3 ml of sterile distilled water were added twice a week to keep cultures wet and amoebic growth was observed daily by microscope examination for a wet mount slide for 4 week. The identity of *Balamuthia mandrillaris* was confirmed, after morphological characterization, genetically by conventional PCR using a set of *B. mandrillaris* specific two primers designed by Booton *et al.* (2003 a) : 5Balspec 16S (5'-CGCATGTATGAAGAAGACCA-3) and 3Balspec 16S (5'-TTACCTATATAATTGTCGATACCA-3). (manufactured by Alpha DNA).

Genomic DNA from cell culture of *Balamuthia mandrillaris* were extracted by using gSYNC™ DNA Extraction kit, Geneaid, Korea, and done according to company instructions. The PCR product yield was a 1075 bp from mitochondrial small-subunit-rRNA genes in *B. mandrillaris* according to the following protocol :

Initial denaturation 10 min at 95C⁰ , 35 cycle of 35 sec at 95C⁰ , 50 sec at 49 C⁰ and 40 sec at 72 C⁰ followed by 10 min final extension at 72 C⁰ ,PCR product was electrophoresed on 1.5% agarose gel and visualized by UV.

RESULTS

B. mandrillaris cyst and trophozoite were observed in 12(16%) out of 75 samples from different environmental sources , the highest occurrence was in tanky water was 2 (50%) and no occurrence was in both tap water ,marshes water , potato soil and wild mice waste . Among the 12 suspected environmental isolates that confirmed by the presence of distinct amoeba cyst or both trophozoites and cyst , only 9 (12%)were positive after the PCR method using *Balamuthia mandrillaris* specific primer Balspec 16S forward and reverse that amplified a portion of mitochondrial rRNA gene yield a 1075 bp product . Table (1) Fig (1) .

Table (1) : Occurrence of *Balamuthia mandrillaris* in environmental samples obtained by microscopic and molecular examination .

Type of Sample	No. sample EX. By microscope	Microscopic Positive samples		No. samples Ex. By PCR	PCR positive samples	
		No.	%		No.	%
River water	8	2	25	2	1	12.5
Tap water	5	0	0	0	0	0
Tanky water	4	2	50	2	2	50
Stagnant water	4	1	25	1	1	25
Marshes water	5	0	0	0	0	0
Air conditioner	4	1	25	1	1	25
Soil	23	4	17.39	4	2	8.69
Potato soil	4	0	0	0	0	0
Lizard waste	7	1	14.28	1	1	14.2
Birds waste	5	1	20	1	1	20
Mice waste	6	0	0	0	0	0
Total	75	12	16	12	9	12

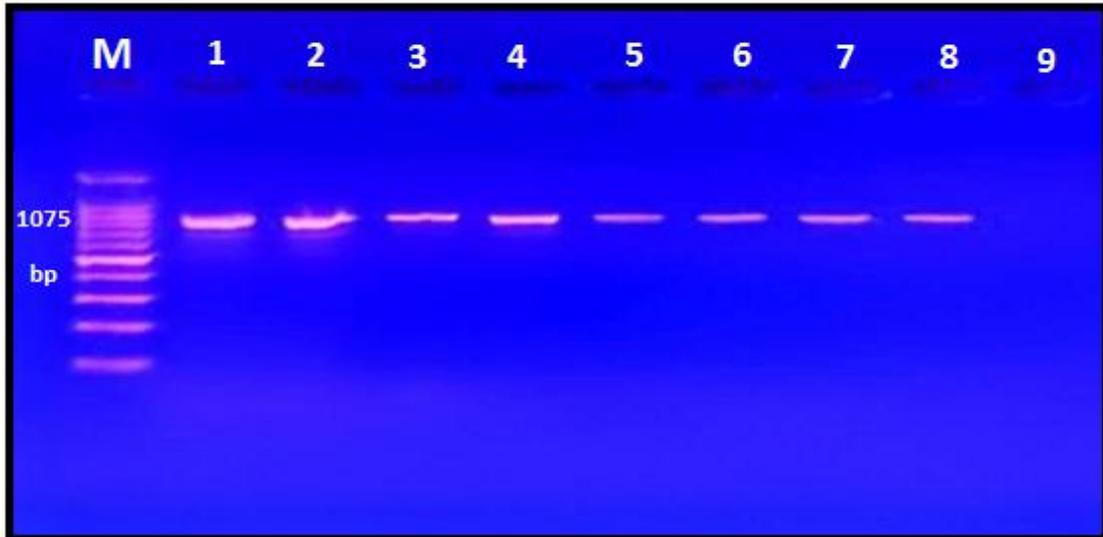


Fig. (1) : Agarose gel electrophoresis image that show the PCR product analysis of 16S ribosomal RNA gene from genomic DNA of *Balamuthia mandrillaris* . from environmental samples : Where M: Marker (2000- 100 bp) lance (1-8) positive samples and lance (9) negative samples at 1075 bp .

Our current study showed *B. mandrillaris* has two stage a vegetative trophozoite stage and dormant cyst stage . The trophozoite stage approximately 35-50 micron in diameter that contain a single nucleus although binucleated form have been observed . It has finger like pseudopodia , transparent ectoplasm and granular endoplasm that contain food and contractile vacuoles . *Balamuthia* trophozoite showed an ability to produce pseudopodia from any part of the amoebic body , trophozoite variable in shape depending on cultivation condition and age of culture Fig. (2)

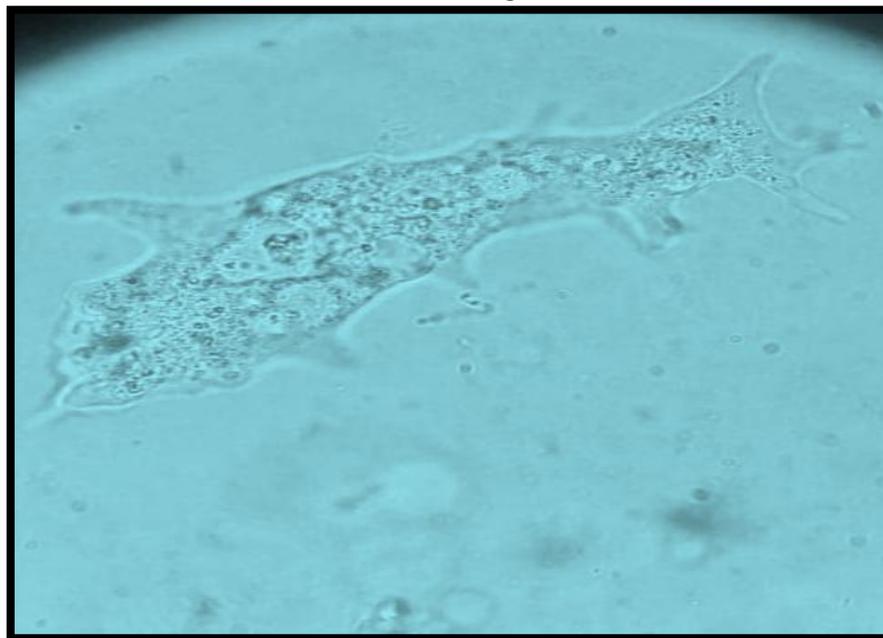


Fig (2) *B. mandrillaris* trophozoite (unstained) 100X

The cysts of *B. mandrillaris* are spherical uninucleus and approximately 12-25 micron surrounded by a double layer membrane , an inner wall that is endocyst and an outer

wrinkled wall that is ectocyst . Both layer are separated with a middle layer composed of amorphous material . Fig. (3) The ectocyst had no pores like *Acanthamoeba* and endocyst without arms ,rounded do not reach the outer edge of outer cyst like *Acanthamoeba* . Those features are helpful in morphological recognition between *Balamuthia* and some species of *Acanthamoeba* .

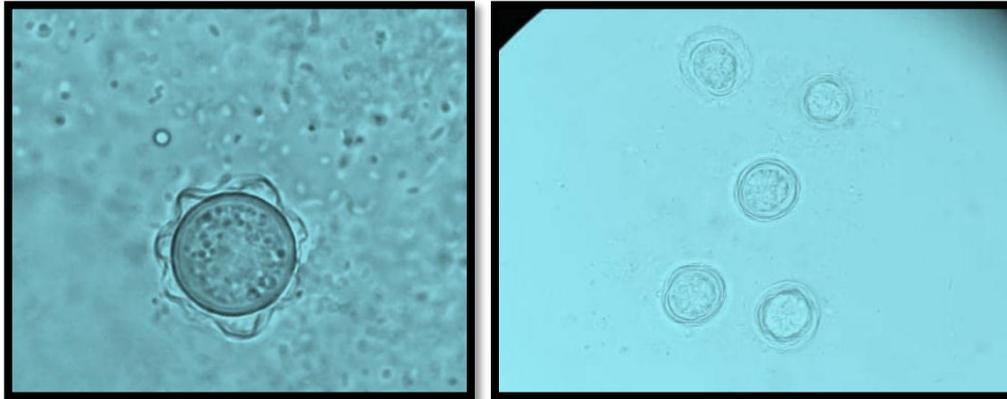


Fig (3) *B. mandrillaris* cyst (unstained)100X

DISSCUSION

B. mandrillaris is a free- living amoeba that lives in soil and water near human settlements (Kanako *et al.*, 2018) . It was supposed to occupy the same ecological habitats of *Acantamoeba* and found naturally in soil (Booton *et al.*, 2003b) . *B. mandrillaris* has been reported as the causative agent of fatal Balamuthia amoebic encephalitis (BAE) (Niyiyati *et al.*, 2015) . Nevertheless environmental isolation of *B. mandrillaris* is rare event and strains of this amoebic species have been isolated from soil and dust sources only in several previous report. The current study is first in Thi- Qar province , that screening environmental detection and isolation of *B. mandrillaris* .In the current study *B. mandrillaris* was observed and diagnosed in 12 (16%) of 75 environmental samples by cultivation and microscopic examination , *B. mandrillaris* was isolated from soil , water and animals waste . PCR using species specific primer was done to confirm the identity ,only 9(12%) samples were positive . There are a number studies that confirmed the presence of *B. mandrillaris* in environmental samples , two cases isolated from the soil in flowerpots in the USA (Schuster *et al.*, 2003 ; Yagi *et al.*, 2005) , two from Iran (One from soil and one from city dust) (Niyiyati *et al.*, 2009; 2016) ,one from well water in Guinea – Bissau (Baquero *et al.*, 2014) ,one from Peruvian soil (Cabello- Vilchez *et al.*, 2014) , one from water in Mexico (Lares – Jimenez *et al.*, 2014) ,one from a mud bath in Jamaica (Todd *et al.*, 2015) , one from soil in the northern region of Japan (Kanako *et al.*, 2018) and one from Shatt AL-Arab the main river in Basrah south of Iraq (Moker & Muslim 2018).

The presence of *B. mandrillaris* in soil and animals waste may refer to a need for organic resources this is agree with first isolation of *B. mandrillaris* from plants pots (Schuster *et al.*, 2003) , where organic fertilizer usually used . This presence could be in associated with the abundant of bacteria and other organismsin such environment that *Balamuthia* could feed on.

The isolation of *B. mandrillaris* from water samples in this study may reveal pollution with organic residues that encouraged their growth and its feeds on other amoebas , the water

samples were poured directly into NN-agar medium without filtration or concentration this reflect the high abundance of *B. mandrillaris* in water samples .

Morphological identification was confirm by PCR method, study PCR is more sensitive technique than direct microscopy of culture , but the use of both PCR and culture method is suggested for environmental samples to gain more complete results of the real presence of *Balamuthia mandrillaris* .

B. mandrillaris may serve as a biological host as well as a transmission vector for some pathogenic bacteria (*Matin et al.* , 2008) , the presence of *B. mandrillaris* can be considered a health hazard , as *B. mandrillaris* is an opportunistic amoeba and also may harbor some pathogenic bacteria .

REFERENCES

- Booton , G. C. ; Carmichae , J. ; Visvesvara , G. ; Byers ,T. and Fuerst , P. (2003)a.** Identification of *Balamuthia mandrillaris* by PCR assay using the mitochondrial 16s rRNA gene as a target . *Journal of clinical microbiology* , 41(1) : 453-455.
- Booton , G. C. ; Schuster , F. ; Carmichael , J. R. ; Fuersta , P. A. and Byers , T. J. (2003)b** *Balamuthia mandrillaris* : identification of clinical and environmental isolate using genus – specific PCR .*J. Eukaryot. Microbiol.* Pp.508-509.
- Baquero, R.A. ; Reyes-Batlle , M. ; Nicola , G.G. ; Martin- Navarro, C.M. et al. (2014).** Presence of potentially pathogenic free- living amoebae strains from well water samples in Guinea – Bissau . *Pathog. Glob. Health.* 108(4) :206- 211.
- Cabello- Vilchez, A.M. ; Reyes –Batlle , M. ;Montalban-Sandoval , E. ;Martin- Navarro, C.M. ;et al.(2014).** The isolation of *Balamuthia mandrillaris* from environmental source from Peru. *Parasitol. Res.* 113(7) : 2509-2513.
- Finnine , P. ; Visvsevara , G. ; Campbell, B. ; Fry , D. and Gasser , R. (2007)** Multifocal *Balamuthia mandrillaris* infection in a dog in Australia . *Parasitol. Res.* , vol. 100(2) :423-46
- Jennifer , R.C. ; Janet , L. ; Hannah , N. ;Sarah , A. C. ; Carol , G. ; Melanie , M. ;Raghuveer , P. ;Jonathan , S. Y. ; Ibne K.A. and Sharon , I. R. (2019).** The epidemiology and clinical features of *Balamuthia mandrillaris* disease in the United States , 1974-2016 . *Clinical Infectious Disease* , vol. 68 (11) :pp1815-1822 .
- Kanako , Y. ; Hiroki , A. ; Yamato , S. ; Kazuki , K. ; Kosuke , K. ; Koichi , I. and Takashi , I. (2018)** . First report of the isolation of *Balamuthia mandrillaris* in the northern region of Japan . *Parasitol . Research* ,117: 2895-2900.
- Lokhande , S. ; More , B. ; Nikam , S. ; Sontakke , T. ;Bandra , V. and Bansode , V. (2015)** . Morphological studies on two rare water amoeba *Balamuthia mandrillaris* and *Vannella mirodes* from Godavari basin at Gangapur and Vaijapur . *Int. J. Recent . Sci. Res.*, vol. 6(6): 4386- 4388.
- Lares –Jimenez , L.F. ; Booton ,G.C. ; Lares-Villa , F. ; Velazquez – Contreas , C.A. and Fuerst ,P.A. (2014)** . Genetic analysis among environmental strains of *Balamuthia mandrillaris* recovered from an artificial lagoon and from soil in Sonora , Mexico. *Exp. Parasitol.* 145(S):57-61.
- Matin ; Siddiqui , R. ; Jayasekera , S. and Khan , N.A. (2008)** . Increasing importance of *Balamuthia mandrillaris* . *Clin. Microbiol. Rev.*, vol. 21(3): 435-448.

Moker , H.M. and Muslim ,A.M. (2018). Isolation of *Balamuthia mandrillaris* (Free-living amoeba) from Shatt AL-Arab River in Basrah , South of Iraq . International Journal of Sciences ,7(1) :90-93.

MMWR.(2010). Morbidity mortality weekly report . *Balamuthia mandrillaris* Transmitted Through Organ Transplantation , Mississippi , 2009 . 59 (36) : 1165-1170.

Niyyati , M. ; Karamati , S. A.; Lorenzo- Morales , J. and Lasjerdi , Z. (2015). Isolation of *Balamuthia mandrillaris* from soil samples in north – western Iran . Parastol. Res.

Niyyati , M. ; Lorenzo- Morales , J. ; Rezaeian , M. ; Martin – Navarro, C. M. ; Hagi, A. M. ;Maclver , S.K. and Valladares , B. (2009). Isolation of *Balamuthia mandrillaris* from urban dust , free of known infectious involvement . Parasitol. Res. 106 (1) : 279-281 .

Niyyati , M. ;Karamati , S. A. ; Lorenzo-Morales , J. and Lasjerdi , Z. (2016). Isolation of *Balamuthia mandrillaris* from soil samples in North-Western Iran . Parasitol. Res. 115(2) : 541-545.

Schuster , F. L. (2002) . "Cultivation of pathogenic and opportunistic free-living amoeba .Clinical Microbiology Reviews , Vol. 15 ,no.3 ,pp. 342- 354.

Schuster, F.L.and Visvesvara, G.S.(2008)."*Balamuthia mandrillaris* " in Emerging protozoan Pathogens , N.A.Khan , Ed., pp. 71- 118 .Taylor &Francis , New York , NY , USA

Schuster , F.L. ; Dunnebacke , T. H. ; Booton ,G.C. ; Yagi , S. ; Kohlmeier , Cc. k. ; et al. (2003). Environmental isolation of *Balamuthia mandrillaris* associated with a case of amoebic encephalitis . J. Clin. Microbiol. , 41(7) : 3175 – 3180 .

Siddiqui , R. and Khan , N.A.(2015).*Balamuthia mandrillaris* :Morpholgy , Biology and virulence . Trop. Parasitol. 5 (1) :15-22 .

Todd ,C.D. ; Reyes – Batlle ,M.; Pinero, J.E. ; Martinez- Carretero, E. ;Valladares ,B. ;Lindo , J.F. and Lorenzo- Morales , J. (2015) . *Balamuthia mandrillaris* therapeutic mud bath in Jamaica . Epidemiol. Infect. 143:2245-2248.

Visvesvara ,G.S. ; Moura , H. and Shuster, F.L. (2007). Pathogenic and opportunistic free-living amoebae , *Acanthamoeba spp.* ,*Balamuthia mandrillaris* , *Naegleria fowleri* and *Sappina diploidea* . FEMS Immunoi. Med. Microbiol . 50,(1) 1-26.

Yagi , S. ;Booton , G. ; Visvesvara , G. and Schuster , F. (2005) . Detection of *Balamuthia mandrillaris* 16s rRNA gene DNA in clinical specimens by PCR . J. Clin . Microbiol. 43(7) 3192- 3197 .